



International journal of Advanced Biological and Biomedical Research

Volume 1, Issue 11, 2013: 1315-1319



# Rubia tinctorum L. (Rubiaceae) or Madder as one of the living color to dyeing wool

# Hamze Esalat Nejad<sup>1</sup>, Ahmad Esalat Nejad<sup>2</sup>

<sup>1</sup> Graduate of Master Textile Engineering, branch Tehran, Islamic Azad University, Tehran, Iran <sup>2</sup> Specializing in the affairs herbal and industrial dyeing crude materials for exquisite carpet weaving, Kashan, Iran

## ABSTRACT

The medicinal part of *Rubia tinctorum* is the dried root. The small yellowishgreen flowers are in loose, leafy, long-peduncled terminal or auxiliary cymes. The margin of the calyx is indistinct, 4- to 5-sectioned and has a tip that is curved inward. There are five stamens and an inferior ovary. The fruit is a black, peasized glabrous, smooth drupe containing two seeds. The perennial plant grows to a height of 60 to 100 cm. The pencil thick rhizome creeps widely underground. The stem is quadrangular with backward turning prickles at the edges.

Key words: Rubia tinctorum, Madder, dyeing wool, red color

## INTRODUCTION

The stems are at times so thin that they are more descendent than erect. The leaves are in whorls, in fours below, in sixes above. They are oblong to lanceolate with one rib and protrudingly reticulate beneath. The plant is indigenous to southern Europe, western Asia and North Africa, and is cultivated elsewhere (Medical Economics Co., 2000). Shape of madder showed in figure1.



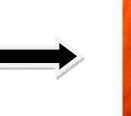




Figure 1: madder plant use to provide red colors

### **Chemical constituents**

Compounds found in Rubia tinctorum include purpurin (oxyalizarin; 1,2,4-trihydroxyanthraquinone), mollugin (6-hydroxy-2,2-dimethyl-2H-naphtho[1,2-b]pyran-5- carboxylic acid, methyl ester), 1-hydroxy-2-methylanthraquinone, 2-ethoxymethylanthraquinone, rubiadin (1,3-dihydroxy-2-methylanthraquinone), 1,3-dihydroxyanthraquinone, hydroxy-2-methylanthraquinone, lucidin(1,3-dihydroxy-7-2hydroxymethylanthraquinone), 1- methoxymethylanthraquinone, 2,6-dihydroxyanthraquinone, lucidin3o-primeveroside[6-(β-d-xylosido)-d-glucoside], alizarin(1,2-dihydroxyanthraquinone), lucidin-O-ethyl 1-hydroxy-2-hydroxymethylanthraquinone 3-glucoside, 2-hydroxymethylanthraquinone ether. 3-3,8-dihydroxy-2-hydroxymethylanthraquinone3-glucoside, ruberythric acid glucoside, (alizarin primeveroside; alizarin glycoside), quinizarin and iridoid asperuloside (Schneider et al., 1979; Kawasaki et al., 1992; Derksen et al., 1998; El-Emary & Backheet, 1998; Marczylo et al., 2000). A number of compounds have been characterized from the roots of R. tinctorum (the source of commercial madder colour) by various analytical methods. Among these compounds are alizarin, ruberythric acid, purpurin, lucidin, rubiadin, mollugin, 1-hydroxy-2- methylanthraquinone, tectoquinone (2-methylanthraquinone), (1,3-dihydroxy-2-anthraquinonecarboxaldehyde),1-hydroxy-2methoxyanthraquinone, nordamnacanthal 1,3-dihydroxy-2-ethoxymethylanthraquinone, scopoletin (7-hydroxy-6-methoxycoumarin) and the glucosides and/or the primeverosides of these compounds (Kawasaki et al., 1992; Westendorf et al., 1998; Medical Economics Co., 2000). The majority of the anthraquinones present in the plant itself or in plant extracts are glycosides (Blömeke et al., 1992; Westendorf et al., 1998).

#### Use

The roots of R. tinctorum contain a red coloring matter which is used for dyeing. Additionally, extracts from R. tinctorum are used for the treatment of kidney and bladder stones (Westendorf et al., 1990; Blömeke et al., 1992). Plants containing 1-hydroxyanthraquinone have been widely used for pharmaceutical purposes such as treatment of kidney and bladder stones, as a laxative mixture, and as a mild sedative (Brown & Brown, 1976; Mori et al., 1990; Wang et al., 1992). Madder root has reportedly also been used medicinally for menstrual and urinary disorders (Medical Economics Co., 1998, 2000).



The roots of M. officinalis have been used as a Chinese natural medicine for tonic and analgesic purposes. Anthraquinone glycosides are the active principles of plant-derived laxatives such as senna, cascara, frangula, rhubarb and aloe. These five plant-derived laxative substances are not included in the present review, because there are no published reports on their potential carcinogenicity. 1,8-Dihydroxyanthraquinone, the aglycone moiety of the laxative ingredient of senna, was formerly marketed as a laxative under the trade name Dantron<sup>®</sup>, but human drug products containing Dantron<sup>®</sup> (see IARC, 1990) were withdrawn from commerce in the United States in 1987 after it was shown to cause intestinal tumors in experimental animals.

#### Steps of coloring with natural coloring

For a long time now she has been niggled by the generally accepted stories about steaming dye vats. Unimpressed by rug lore, it seemed to her most unlikely that primitive tribes people would have resorted to such processes. If her feelings were reasonable, then it had to follow that it must be possible to achieve satisfactory results with water at "domestic" temperature. The countries in which rugs are traditionally made have a very warm climate, so a vat left out in the sun would certainly become pretty hot. Admittedly the process would be slower, but time was never a critical matter for primitive peoples who, in any case, were generally limited to measuring this in days and in lunar months. so variety of providing colors shown in figure 2.

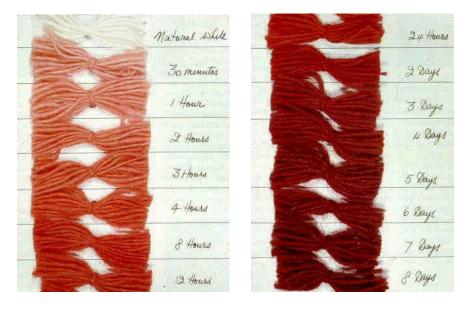


Figure 2. Providing colors after different steps of dyeing

With this in mind, she experimented with various mordant strengths, various concentrations of dye stuff, various periods of immersion, and various temperatures. Using madder in this way she achieved an incredible range of colors from the softest "decorator" pinks to the deep masculine reds so often found in Turcoman rugs. But her delight arose from the fact that her results could be consistently replicated. Thanks to modern photography and an excellent PC with a DTP program, she has crystallized these into picture and print as you can see. She has provided me with a recipe for cool dyeing with madder root, a card showing the deepening color in 15 individual swatches of natural white wool immersed for increasing periods from half an hour to eight days, and a color photograph showing the consequences of heat to swatches immersed for four different periods of time and at three different temperatures. Published treatises on dyeing with madder root have all warned the reader of the complexity and critical nature of the process, requiring the wool to be heated to temperatures likely to cause it to felt. The purpose of this experiment is to discover if it is possible to dye wool to deep brown-red shades without heating the dye pot to temperatures detrimental to the fiber.

## **Experiment 1**

### Esalat Nejad

Wool was placed in a cold hard water bath containing Alum mordant in the proportion of 25% Alum to wool by weight and left to soak for 30 days. After removal from the vessel, the wool was rinsed in cold water.

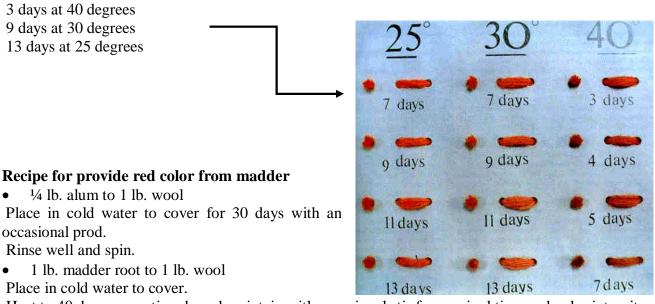
The wool was then placed in a vessel with an equal weight of chopped madder root and water to cover. The vessel was brought up to a temperature of 40 degrees centigrade in this case by simply placing the vessel on the back of an Aga between the hot plates and further insulating the vessel with the appropriate number of pot



holders underneath. Samples of wool were taken from the dye pot on the third, fourth, fifth, and seventh days. The results are shown in

#### **Experiments 2 and 3**

If it is possible to achieve such intensity of color within seven days at a temperature of 40 degrees, I was curious to know just how successful it would be at colder temperatures. Two vessels were prepared, the first at 30 degrees centigrade, a temperature which can be maintained by placing the pot near a radiator, for example. The other pot was left a room temperature, which was a constant 25 degrees centigrade. The results of these experiments show that given enough time, very intense shades of red and red-brown are achievable without any effort or skill involved in the process. The time comparison to achieve a similar shade:



Heat to 40 degrees centigrade and maintain with occasional stir for required time and color intensity.

## REFERENCES

Blömeke, B., Poginsky, B., Schmutte, C., Marquardt, H. & Westendorf, J. (1992) Formation of genotoxic metabolites from anthraquinone glycosides present in Rubia tinctorum L. Mutat. Res., 265, 263–272

Blumenthal, M., Busse, W.R., Goldberg, A., Gruenwald, J., Hall, T., Riggins, C.W. & Rister, R.S., eds (1998) The Complete German Commission E Monographs: Therapeutic Guide to Herbal Medicines, Austin, TX/ Boston, MA, American Botanical Council/Integrative Medicine Communications.

Derksen, G.C.H., Van Beek, T.A., De Groot, A. & Capelle, A. (1998) High-performance liquid chromatographic method for the analysis of anthraquinone glycosides and aglycones in madder root (Rubia tinctorum L.). J. Chromatogr. A., 816, 277–281.

El-Emary, N.A. & Backheet, E.Y. (1998) Three hydroxymethylanthraquinone glycosides from Rubia tinctorum. Phytochemistry, 49, 277–279

Felter, H.W. & Lloyd, J.U. (2002) King's American Dispensatory. Rubia-Madder (http://ftp.oit. unc.edu/herbmed/eclectic/kings/rubia.html).

Fujita, M., Furuya, T. & Matsuo, M. (1961) Studies on the metabolism of naturally occurring anthraquinones. I. The metabolism of 1-hydroxyanthraquinone and 2-hydroxyanthraquinone. Chem. pharm. Bull., 9, 962–966.

Krizsan, K., Szokan, G., Toth, Z.A., Hollosy, F., Laszlo, M. & Khlafulla, A. (1996) HPLC analysis of anthraquinone derivatives in madder root (Rubia tinctorum) and its cell cultures. J. liq. Chromatogr. relat. Technol., 19, 2295–2314.

Mori, H., Mori, Y., Tanaka, T., Yoshimi, N., Sugie, S., Kawamori, T. & Narisawa, T. (1992) Cell kinetic analysis of the mucosal epithelium and assay of ornithine decarboxylase activity during the process of 1-hydroxyanthraquinone-induced large bowel carcinogenesis in rats. Carcinogenesis, 13, 2217–2220.

Murti, V.V.S., Seshadri, T.R. & Sivakumaran, S. (1972) Chemical components of Rubia iberica. Indian J. Chem., 10, 246–247.

Poginsky, B., Westendorf, J., Blömeke, B., Marquardt, H., Hewer, A., Grover, P.L. & Phillips, D.H. (1991) Evaluation of DNA-binding activity of hydroxyanthraquinones occurring in Rubia tinctorum L. Carcinogenesis, 12, 1265–1271.

Westendorf, J., Poginsky, B., Marquardt, H., Groth, G. & Marquardt, H. (1988) The genotoxicity of lucidin, a natural component of Rubia tinctorum L., and lucidinethylether, a component of ethanolic Rubia extracts. Cell Biol. Toxicol., 4, 225–239.

Westendorf, J., Marquardt, H., Poginsky, B., Dominiak, M., Schmidt, J. & Marquardt, H. (1990) Genotoxicity of naturally occurring hydroxyanthraquinones. Mutat. Res., 240, 1–12.

Westendorf, J., Pfau, W. & Schulte, A. (1998) Carcinogenicity and DNA adduct formation observed in ACI rats after long-term treatment with madder root, Rubia tinctorum L. Carcinogenesis, 19, 2163–2168.

Yang, Y.J., Shu, H.Y. & Min, Z.D. (1992) Anthraquinones isolated from Morinda officinalis and Damnacanthus indicus. Yao Xue Xue Bao (Acta pharm. sin.), 27, 358–364