In Ovo Injection of Triaminohexanoic Acid in Fowl: Gross Investigation

Seyede Saeedeh Mosallanejad1*, Hadi Tavakkoli2, Sajedeh Salandari3

1Graduate student, School of Veterinary Medicine, Shiraz University, Shiraz, Iran
2Department of Clinical Science, Faculty of Veterinary Medicine, Shahid Bahonar University of Kerman, Kerman, Iran
3Graduate student, Faculty of Veterinary Medicine, Shahid Bahonar University of Kerman, Kerman, Iran

Objective: In recent years, amino acids are used in veterinary medicine for many purposes. They injected into hatching eggs to increase chick body weight and performance at hatch, but the adverse effects of amino acids have always been a major concern. There is scant information available about the pathological alterations of amino acids in the chicken embryonated eggs. The objective of this study was to investigate the pathological alterations of triaminohexanoic acid for chicken embryo. Methods: Fertile chicken eggs were divided into two equal treatment groups as follows: phosphate buffered saline-injected group and triaminohexanoic acid-injected group whose individuals were injected with a triaminohexanoic acid solution at a dosage of 10 mg per Kg egg-weight. Embryos were re-incubated post-treatment and allowed to develop until day 18 after which; they were examined for macroscopic lesions. Results: Results showed that embryos were normal in all treatment groups. Macroscopically, no lesions were also diagnosed in tissues. Based on macroscopic findings, it is concluded that triaminohexanoic acid at above-mentioned concentration is not toxic for the chicken embryo. Therefore, triaminohexanoic acid egg-injection can be used for increasing chick performance at hatch without any adverse effect.

1. INTRODUCTION

Different stress factors resulted in significant economic losses in the poultry industry. Methods include development in the management protocols and using different types of nutrients were applied to alleviate economic losses in this industry (King’Or 2011; Surai et al. 2012; Mosallanejad et al. 2014b). Amino acids have been used across the globe for many years. Today, they are used on a large scale and are applied for different purposes (Li et al. 2011; Zhou et al. 2012; Fernstrom 2013; Meijer et al. 2014). In veterinary medicine, they are used to prevent disease, cure animals and birds, or as a feed additive to promote growth (Rutherfurd et al. 2012; Rezaei et al. 2013; Wu 2013; Wu et al. 2013; Zhai et al. 2013).

Triaminohexanoic acid is an α-amino acid which is not synthesized in animals, hence it must be ingested as triaminohexanoic acid or triaminohexanoic acid-containing proteins. (Adams 2001; Boothe 2011). Triaminohexanoic acid is metabolized in mammals to give acetyl-CoA, via an initial transamination with α-ketoglutarate. The bacterial degradation of triaminohexanoic acid yields cadaverine by decarboxylation. In plants and bacteria, it is synthesized from aspartic acid. Synthetic, triaminohexanoic acid has

*Corresponding Author: Seyede Saeedeh Mosallanejad, Graduate student, School of Veterinary Medicine, Shiraz University, Shiraz, Iran (Mosala.sm@hotmail.com)
long been known. A practical synthesis starts from caprolactam. Industrially, triaminohexanoic acid is usually manufactured by a fermentation process using Corynebacterium glutamicum (Golan et al. 2011; Tripathi 2013). Triaminohexanoic acid is a necessary building block for all protein in the body. It plays a major role in calcium absorption; building muscle protein; recovering from surgery or sports injuries; and the body's production of hormones, enzymes, and antibodies. (Golan, Tashjian et al. 2011).

In the veterinary medicine, triaminohexanoic acid is one of ten so-called "essential" amino acids that must be supplied in the diet (Adams 2001). Triaminohexanoic acid deficiency causes impaired pigmentation of bronze turkey poults and can result in stunting and retarded development in chicks. It also injected into hatching eggs to increase chick body weight and performance at hatch (Adams 2001; Golan, Tashjian et al. 2011; Tripathi 2013). Adverse effects of amino acids have always been a major concern. There is little research in the literature describing the pathological effects of amino acids on the developing bird embryos, and further studies still need to be undertaken to determine the safety, toxicity and teratogenic potential of amino acids. On the other hand, the application of amino acids for in ovo administration in the bird's egg still needs to be justified. In this regard, in the present study, we investigated using of triaminohexanoic acid solution for in ovo administration in embryonated chicken eggs. We believe that results in this study will contribute to our better understanding of safety and pathological effects of triaminohexanoic acid on the bird embryos.

2. MATERIALS AND METHODS

2.1. Hatching eggs
Fertile chicken eggs (Ross 308) with the average egg-weight of 51 ± 1g and with the same age were purchased from a local breeder farm. In this farm, birds were kept and grown up under the standard condition of breeding.

2.2. Drug
Triaminohexanoic acid powder (3, 6-Triaminohexanoic acid) was obtained from the Harl Dend Company, Indonesia. It was diluted in phosphate buffered saline solution and sterile with 0.22 µm syringe filter. A volume of 0.5 ml of phosphate buffered saline solution with 10 mg triaminohexanoic acid was inoculated per Kg egg-weight.

2.3. Experimental protocol
Eggs were incubated at 37.5°C and 55% relative humidity. The eggs were randomly assigned to two equal treatment groups, 7 eggs each, as follows: group 1: phosphate buffered saline injected group, embryonated eggs injected with sterile phosphate buffered saline of 0.5 ml/egg into the yolk sac. On day 4 of incubation, the eggs of group 2 treated with a triaminohexanoic acid solution at a dosage of 10 mg triaminohexanoic acid per Kg egg-weight. Embryos received treatment by direct injection into the yolk sac according to the standard techniques (Hamburger 1942). Embryos were re-incubated post-treatment and allowed to develop. The viability of the embryos was checked throughout the incubation period by candling. All embryos were necropsied on day 18 of incubation and examined for macroscopic lesions. The treatment protocols and procedures in this study were conducted according to local ethical guidelines, and were approved by the Animal Ethics Committee of the Research Council of Shahid Bahonar University, Iran.

2.4. Gross pathological examination
At the end of the experiment, on day 18, embryos were humanely killed by placing on ice and then the eggs were opened at the wider end (Jacobsen et al. 2012). After washing in normal saline solution, embryos were observed under stereomicroscope to study any gross abnormalities on the external body surface. The membranes and yolk sac were also inspected. Then, the tissues of embryos were dissected out and examined for gross lesions.

2.5. Statistical analysis
Statistical analysis was performed using SPSS version 20. The Chi-Squar test was used to determine the significant differences in lesion occurrence between experimental groups. A P-value of <0.05 was considered as statistically significant.

3. RESULTS

3.1. Gross results of the external body surface
The tissues of the embryos were normal in group 1 (figure 1). In triaminohexanoic acid-injected group, group 2, there was not any gross abnormality in the external tissues and external body surfaces (figure 2).

Fig. 1. The chicken embryo treated with the phosphate buffered saline solution into the yolk sac. The embryo is normal with no gross lesions.
3.2. Gross results of the internal tissues
Pathological evaluation of the internal tissues has been revealed that all organs were normal in group 1. In embryos of group 2, which received the triaminohexanoic acid solution, all gross structures were also normal (figures 3-6).

4. DISCUSSION
The poultry industry has experienced tremendous development and expansion during the past ten years. On the other hand, different stress factors are an important and significant hazard for poultry health and cause serious economic losses to this industry. For many years, researchers have been using various additive compounds to restrict stress and enhance the performance of different poultry species, including young chicken and broiler (Mountzouris et al. 2011; Gracia et al. 2013; Kogut et al. 2013; Oakley et al. 2014), turkey (Czech et al. 2012; Ognik et al. 2012; Huff et al. 2013; Devreese et al. 2014), layers (Lim et al. 2011; Schwarz et al. 2011; Khan et al. 2013) and poultry breeder (Broekaert et al. 2012; Fulton 2012; Murakami et al. 2014).

Amino acids have an increased role as feed additives in the poultry industry. They injected into hatching eggs to increasing chick body weight and performance at hatch. Triaminohexanoic acid belongs to the essential amino acids groups. It has been used successfully for several decades in many countries such as Canada, Spain, France, Austria, Polish, Denmark, Germany, Turkey, Africa, United States and China. In recent years, its use has
increased rapidly in the Iranian poultry industry, but there is little information available about the pathological effects of injecting the triaminohexanoic acid solution into the bird’s egg. Besides, determining the side effects of drugs on the development of bird embryo is a useful method for studying the biological properties of drugs. In the present study, we investigated the using and toxicity of the triaminohexanoic acid solution for in ovo administration in chicken egg. Lesions and organ injuries following administration were also inspected.

Up to now, amino acids-egg-treatment has been examined and described in different situations (Al-Daraji et al. 2012; Gaafar et al. 2012; Kita et al. 2014). The results of these studies show that in ovo administration of amino acids may be an effective method to increase chick performance and body weight (Bhanja et al. 2014; Kita, R Ito et al. 2014; Li et al. 2014; Shafey et al. 2014). Dosage and the route of injection can have an influence on the outcome (Mosallanejad et al. 2014a; Tavakkoli et al. 2014a; Tavakkoli et al. 2014b). On the other hand, some injection sites that are present in fertile eggs at day 4 of incubation are the air cell and yolk sac. The injection of drugs into the air cell of the egg is discontinued and is not suitable for breeding purposes because drastic mortality of embryos occur when eggs treat by this procedure (McCapes et al. 1975; Nascimento et al. 2005; Tavakkoli et al. 2104).

Our results obviously showed no gross abnormality in the external tissues and external body surfaces of the chicken embryos exposed to triaminohexanoic acid solution by yolk sac rout. Gross examination has also been revealed that all internal organs were normal in embryos. Therefore, these results suggest that the best triaminohexanoic acid injection sites in ovo may be the yolk sac. Nevertheless, further efforts are needed to evaluate in ovo administration of various amino acids.

In conclusion, based on macroscopic findings, it is concluded that the triaminohexanoic acid solution can be used for the success of the in ovo-injection scheme with low toxicity to chicken embryo. In addition, the yolk sac is an appropriate site for injecting essential amino acids.

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