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## **Original Article**

# Effect of Seed Priming on Germination and Seedling Growth of the Caper (*Capparis Spinosa*) Under Drought Stress

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#### ARTICLE INFO

#### ABSTRACT

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**Objective:** To evaluate the effect of chemical stimuli on germination and seedling growth of Caper (*Capparis Spinosa*) under drought stress. **Methods:** Factorial experiment in a completely randomized design with four replicates per treatment were performed. First factor of priming include three levels of acetylsalicylic acid (300, 200 and 100 mg/L), gibberellic acid levels (500, 250 and 125 ppm), ascorbic acid levels (300, 200 and 100 mg/L) and distilled water as a control and the second factor was drought stress levels (0, -0.3, -0.6, -0.9 and -1.2 MPa). **Results:** The results showed that priming increases germination rate, germination percent, root length, seedling length, shoot length and seed vigor index under drought stress. Among of levels used in priming, acetylsalicylic acid 200 mg/L has greatest effect on the germination of plants under drought stress. According to the survey results, priming with salicylic acid compared with other acids have more effect on germination characteristics of Caper under drought stress. **Conclusions:** Thus, using of this method can be useful for improving seed germination characteristics of plants in arid and semi-arid regions.

#### **1.INTRODUCTION**

Plants play an important role in providing food for humans (Abdolshahi et al., 2013; Rad et al. 2013). During their life cycle, plants endure a different of abiotic stresses such as high and low temperatures, drought, salinity, heavy metals, UV and highlight, Allelochemical substances, chemical compounds which can have a profound effect on viability, growth, morphology, and reproduction (Rad et al., 2013; Miri et al., 2013; Rad et al. 2014 a; Rad et al., 2014 b). Water shortage is a major problem for agricultural production in the world. Iran with an average of 240 mm of rainfall is located in arid regions of the world (Kazemepoor and Orvin, 1998). Increasing drought decreases germination rates that it can be as an adaptation mechanism of native plant to drought condition and amount of reduction depends on plant species and the drought intensity (Maraghni et al., 2010; Zeng et al., 2010). One way of increasing germination in stress condition is priming method (Maraghni et al., 2010). Seed priming is the process of regulating germination by managing the temperature and seed moisture content; in order to maximize the seed's potential. Seed priming can improve germination rate, reduce time of germination and seedling emergence and improve plant establishment. There are evidence regarding the use of chemical stimuli in accelerating growth and germination (Imani et al., 2014). Growth

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hormone is normally used for seed priming, including auxin, abscisic acid, polyamines, ethylene, salicylic acid and ascorbic acid (Demiral and Turkan, 2005). Primed seeds with gibberellic acid usually increases the emergence, growth and extensive of root systems. In addition, seed priming with gibberellic acid accelerates flowering, maturity and yield of plants (McDonald, 2000; Senaratna et al., 2000). One way to combat free radicals in seeds is antioxidants vitamins, such as ascorbic acid (vitamin C). Vitamin C is a water-soluble vitamin and eliminates the negative effects of free radicals superoxide and hydroxyl (McDonald, 2000; Rad et al., 2014 c). Seed priming increases the glucose and proline content and can be improves quality of the germination and germination index in dry conditions. However, increasing in drought, germination stopped and it causing serious damage to seedling plants (Sun et al., 2010). Li et al.,(2011), investigated the seed germination of pyrethrum (*Tanacetum cinerariifolium*) and they reported that with increasing salinity, drought and light germination was decreased. Also they are reported that hydropriming improve germination percentage, reduce mean time to germination in all osmotic potential. Their results showed that germination in distilled water increased from 52% to 59% and from 16% to 29% at the highest concentration Salinity level (Li et al., 2011). Fang et al., (2012) showed that under drought conditions in of Salix paragplesia and Hippophae two species rhamnoides, height, base diameter and number of leaves reduced. In this condition, these plants increased root to shoot ratio and therefore they are able to use water in dry conditions (Fang et al., 2012). Environmental stresses such as drought damage plants during germination which damage to their qualitative and quantitative performance and causing weakness and heterogeny in plant growth (Fetri et al., 2014). So, understanding drought tolerance threshold for seed germination is essential and can be an effective step towards increasing the permanent vegetation, soil and water conservation and identify the degree of drought resistance in plants.

#### **2. MATERIALS AND METHODS**

To study the effects of chemical stimulants on seed germination of Caper plant under drought stress, experiments was carried out in the biotechnology Center of University of Zabol, Iran. The seeds Capparis Spinosa was purchased from the Pakan Bazr Company, Isfahan, Iran.. At the first seeds was disinfected by sodium hypochlorite 5% and then were washed several times with distilled water. Then seeds were placed separately in chemical stimuli for 10

h with salicylic acid (100, 200 and 300 mg/l) and 24 h with gibberellic acid (125, 250 and 500 ppm) and 8 h with ascorbic acid (100, 200, and 300 mg/l) at 25 °C. Distilled water was used as control .After the soaking period, all the seeds were washed with distilled water and after that, seeds were placed on sterilized filter paper in 9-cm diameter Petri dishes under different drought level. To prepare different concentrations of drought, polyethylene glycol 6000 were used. To calculate the amount of required PEG for osmotic pressure (Michel and Kaufman, 1973), the following formula was used:

$$\psi s = -(1.18 \times 10-2) C - (1.18 \times 10-4) C_2 + (2.67 \times 10-4) CT + (8.39 \times 10-7) C_2 T$$

 $\psi s$ : Osmotic pressure in terms of Bar

C: The concentration of PEG 6000 g/kg water (g/kg  $H_2O$ )

T: Temperature in terms of centigrade

To prepare zero water potential (control), distilled water were used. Factorial experiment in a completely randomized design with four replications (25 seeds per replicate) in different level of drought (0, -3, -6, -9 and -12Mega - Pascal) was performed. The experiments were carried out in a programmed incubator at  $25\pm2^{\circ}$ C and 16 hours light and 8 hours of darkness. For a period of 15 days, each day germinated seeds that root length was more than 2 mm were counted (Kolsarıcı, 2006). In this experiment, germination percentage, germination rate (Agrawal and Dadlani 1992, ISTA, 2002) and seed vigor index (Agraval, 2005) were calculated based on the following relationships:

$$GP = 100 \times (ni / s)$$

(Agraval, 2005)

 $GR = \sum ni / ti$ 

(Agrawal and Dadlani, 1992)

GP: Germination percentage, S: Total number of seeds, ni Germinated seeds at time ti, GR: Germination rate, ti Number of days after germination. (Ashraf and Foolad, 2005)

Seed vigor index= final germination percentage  $\times$  Seedling length

#### 2.1. Statistical analysis

Statistical analyses were carried out using SPSS 11.5. All data were analyzed by one-way analysis of variance (ANOVA) to determine the treatments effects, and LSD test at 1% level was carried out to determine the statistical significance of the differences between treatments means.

### 3. Results

### 3.1. Percentage of germination

Analysis of variance showed that drought stress and chemical stimuli have significant effect on germination percentage at  $p \le 0.01$ .Comparison of means showed that the use of chemical stimulants increase seed germination of *C. Spinosa* under drought conditions in compared to control. The maximum effect of priming on germination percentage was observed in salicylic acid (200 mg/L), salicylic acid (300 mg/L) and gibberellic acid 125ppm at-0.3 MPa of drought level. The results showed that there is no significant difference between gibberellic acid (125 ppm) and salicylic acid (300 mg/l). In -3.0 MPa of drought level. Lowest germination was observed in the control (Fig. 1).

### 3.2. Germination rate

Chemical stimuli and drought levels have significant effect on the germination rate of *C. Spinosa* at  $p \le 0.01$ .With increasing drought, germination rate significantly decreased. In all levels of drought, priming with salicylic acid (200 mg/l) had the maximum rate of germination. The results showed that various levels of gibberellic acid in compare to ascorbic acid had a greater role in reducing of the negative effects of drought stress (Fig. 2).

### 3.3. Root length

The results of the variance analysis showed that the interaction between chemical stimuli and different levels of drought have significant effect on root length at  $p \le 0.01$ (Table 1). The results showed that root length was reduced with increasing drought stress but all the chemical stimuli increased the root length. The Maximum of root length was recorded in salicylic acid (200 mg/L) and gibberellic acid (125ppm) with 2.15 and 2.16 cm, respectively and the minimum root length was observed in control. Also the results showed that salicylic acid (200 mg/l) increased root length from 0.85 to 2.03 cm under -0.3 MPa of drought level. Furthermore all concentration of salicylic acid increased the root length in 0.6 MPa of drought level in compare to control. Gibberellic acid and ascorbic acid increased the root length at different level of drought (Fig. 3).

### 3.4. Shoot length

Temperature and chemical stimuli had significant effect on shoot length (Table 1).The results showed that shoot length decreased with increasing drought and all chemical stimuli significantly increased the shoot length. The maximum shoot was recorded in salicylic acid (200 mg/L) and salicylic acid (300mg/l) with 8.3 and 8.1 cm, respectively. It seems that among of the three studied acid, salicylic acid has the greatest effect on shoot length (Fig. 4).

# 3.5. Seedling length

According to Table 1 drought stress and chemical stimuli had significant effect on seedling length. For both primed and non-primed seeds, drought stress had significant effect on the seedling length (Fig. 5).

## 3.6. Seed vigor index

The results showed that chemical stimuli increased seed vigor. The maximum and minimum seed vigor indexes were observed in salicylic acid (200 mg/l) and salicylic acid (300 mg/l) with 910 and 828, respectively. The result showed that the acetylsalicylic acid had maximum effect on decreasing of the negative effect of drought stress. After acetylsalicylic acid, gibberellic acid had a positive role in reducing the negative effects of stress (Fig. 6).

# 4. DISCUSSION

Seed germination is negatively affected by drought stress (Rad and Rad, 2013). In this study, drought stress decreased germination percentage and germination. Seeds for germination process must absorb enough water. Soluble material in the medium such as polyethylene glycol reduces water absorption and subsequent seed germination is delayed or stopped (Tobe et al., 2000). Due to osmotic pressure loss under drought stress, water absorption process is disrupted and the alpha-amylase enzyme activity is inhibited (Afza, 2005). With increasing drought stress, root and shoot length decreased. (Bayoumi et al., 2008). Reducing in water absorption by seeds reduces secretion of the stress hormone and enzyme activity and it has negatively effect on seedling growth (Kafe et al., 2005). Also with increasing drought stress, seed vigor decreased. Similar results were reported by Yunial and Nautial (1998) in the study of *Ougeinia dalbergioides* germination. Overall priming improves seed germination under stress and non-stress conditions (Kolsarıcı, 2006, Murungu et al., 2003). Seed priming reduce time of germination and cause to accelerate seed germination. So, it is the reason of superiority of primed seeds in compared with nonprimed seeds (Netondo et al., 2004). Seed priming increases antioxidant enzymes such as glutathione in primed seeds and this enzyme may reduce lipid peroxidation activity during germination and this can lead to increase germination (Hus and Sung, 1997, Baalbaki et al., 1995). Our results showed that the use of salicylic acid significantly increased germination and seedling growth in in C. Spinosa under drought stress. In agreement with the results, earlier reports (Borsani et al., 2002; Demiral and Turkan, 2005) have shown the germination and seedling growth was increased by using of salicylic acid. The mechanism that salicylic acid

increases seed germination is not yet clearly understood. According to Nun et al. (2003), salicylic acid can inhibit the catalase activity. Catalase activity reduction leading to increased hydrogen proxide that it can enhance some germination of seeds. It is possible that 2-6 dihydroxi benzoic acids and salicylic acid stimulate the germination of seed via GA bio-synthesis and act as thermogene inducers (Shah, 2003). Also our results showed that gibberellic acid lead to increasing in all the germination studied traits in comparison with control. One of effective and positive reason for chemical stimuli such as gibberellic acid on the seed germination probably due to hormonal balance and decrease the proportion of growth inhibiting substances such abscisic acid (ABA). Gibberellin (GA) improves the synthesis and secretion of hydrolytic enzymes from aleurone cells. These enzymes then mobilize the endosperm storage reserves that fuel germination and growth (Cirac et al., 2004). In stress conditions, ascorbic acid acts as an effective antioxidant. Ascorbic acid can remove the free radicals in stress conditions especially oxygen radicals and it has important role in stimulating and cell expansion and nutrient absorption in the plant cells under environmental stresses which leads to avoid from the oxidation risk (Smirnoff and Wheeler, 2000, Smirnoff, 1996). Salinity and drought are two major abiotic determinants (Wang et al., 2009) on seed germination and plant growth. High salinity and severe drought could promote the salinization and desertification of land processes which are rapidly increasing on a global scale. For biological reclamation of this region, before sowing of seeds, chemical stimuli particularly gibberellin acid can be used for increasing seed germination percentage and seedling establishment. According to the survey of results, priming with salicylic acid in compared with other acids had a greater impact on Caper germination characteristics under drought stress. Thus, using of this method can be useful for improving seed germination characteristics of plants in arid and semi-arid regions.

### REFERENCES

Abdolshahi A., Majd M. H., S. Rad. J., Taheri M., Shabani A., da Silva J. A. T.. Choice of solvent extraction technique affects fatty acid composition of pistachio (*Pistacia vera* L.) oil. Journal of Food Science and Technology. 2013. 1-6.

Afzal, I., 2005. Seed enhancements to induced salt tolerance in wheat (*Triticum aestivum* L.). Ph.D. Thesis, Agricultural University of Faisalabad, Pakistan, 266 p.

Agraval, R., 2005. Seed technology. Oxford and IBH Publishing Co, 829p.

Agrawal, P.K and Dadlani,M. 1992. Techniques in seed (science and technology). South Asian Publishers, 209p.

Ashraf, M and Foolad, M.R. 2005. Pre sowing seed treatment Ashotgun approach to improve germination,

plant growth, and crop yield under saline and non saline conditions. Advances in agronomy. 88: 223-265.

Baalbaki, R.Z., Zurayk, R.A., M.M. Blelk, M.M. and Tahouk, S.N. 1999. Germination and seedling development of drought tolerant and susceptible wheat under moisture stress. Seed sciences and technology. 27: 291-302.

Bayoumi, T. Y., H. Manal & E.M. Metwali, 2008. Application of physiological and biochemical indices as a screening techniqus for drought tolerance in wheat genotypes. African journal of biotechnology, 14: 2341-2352.

Borsani, O., J. Cuartero., V. Valpuesta & M.A. Botella, 2002. Tomato tosl mutation identifies a gene essential for osmotic tolerance and abscisic acid sensitivity. Plant Journal. 32, 905-914.

Cirac, C., A.K. Ayan & K. Kevseroglu, 2004. The effects of light and some presoaking treatments on germination rate of st. John Worth seeds. Pakistan Journal Biology Science. 7: 182-186.

Demiral, T., & I,Turkan, 2005. Comparative lipid peroxidant, antioxidant systems and praline content in roots of two rice cultivars differing in salt tolerance, Environment Experiment Botany. 53, 247-257.

Esvand, H.R. 2008. Effect of seed priming on seed old physiological quality under drought stress wheat grass. Ph.D. dissertion agriculture. Department of agriculture, university of Tehran.

Fang, J., Wu, F., Yang, w., Zhang, J. and Cia, H. 2012. Effects of drought on the growth and resource use efficiency of two endemic species in an arid ecotone. Acta ecologica sinica, 32(4): 195-201.

Fetri M., Dargahikhoo A., Rajabi M. Effect of drought and salinity tensions on germination and seedling

growth of Common Yarrow (*Achillea millefolium* L.) in laboratory conditions. International journal of Advanced Biological and Biomedical Research. 2(2), 2014: 383-391.

Hus, J.L and Sung,JM. 1997. Antioxidant role of glutatnione associated with accelerated agina and hydration of triploid Warermelon seeds. Physiologa plantarum. 100: 967-974.

Imani A.F., Sardoei A. S., Shahdadneghad M. Effect of H<sub>2</sub>SO<sub>4</sub> on Seed Germination and Viability of *Canna indica* L. Ornamental Plant. International journal of Advanced Biological and Biomedical Research. 2(1), 2014: 223-229.

ISTA., 2002. International rules of seed testing. seed science and Tecnology. 20:53-55.

Kafe,M., A. Nezamy., H. Hoseine & A. Masome, 2005. Physiological effects of drought stress by polyethylene glycol on germination of lentil (*Lens culinaris* Medik.) genotypes. Journal Iran of research Agronomy.3:69-80.

Kaur, S., A.K. Gupta & N.Kaur, 2000. Effect of GA3, kinetin and indole acetic acid on carbohydrate metabolism in chickpea seedlings germination under water stress. Plant growth Regulator. 30: 61-70.

Kazemepoor, N and Orvin,M.J. 1998. Effect of drought and salt stress on germination and seedling growth of Vetch (Vicia sp.). The eight Conference of iran Biology. 800-810.

Kolsarıcı, 2006. Seed treatments to overcome salt and drought stress during germination in sunflower (*Helianthus annuus* L.).Europ. Journal. Agronomy. 24: 291–295.

Li, j., Yin, L.Y., Jongsma, M.A. and Wang, C.Y. 2011. Effects of light, hydropriming and abiotic stress on seed germination, and shoot and root growth of pyrethrum (*Tanacetum cinerariifolium*). Industrial crops and products, 34(3): 1543-1549.

Maraghni , M., M. Gorai & M.Neffati, 2010. Seed germination at different temperatures and water stress levels, and seedling emergence from different depths of *Ziziphus lotus*. South African Journal of Botany. 76(3): 453-459.

Mauromicale, G. & H. Cavallaro, 1997. A comparative study of the effects of different compounds on priming of tomato seed germination under suboptimal temperatures. Seed science and technology. 25: 399 – 408.

McDonald, M.B. 2000. Seed priming. ln 'black, m. g. d. bewley. (eds) . seed technology and lts biological basis. Sheffield academic press, Sheffield, uk, pp, 287 -325.

Michel, B.E & M.R, Kaufman, 1973. The osmotic potential of polyethylene glycol 6000. Plant physiology.51:914-916.

Miri A., Rad J. S., M. S. Rad, da Silva J.A. T.. Allelopathic activity of medical plant, Cardaria draba (*Lepidium draba* L.). Annals of Biological Research, 2013, 4 (6):76-79.

Murungu, F.S., P. Nyamugafata., C. Chiduza., L.J. Clark & W.R, Whalley, 2003. Effects of seed priming aggregate size and soil matric potential on emergence of cotton (*Gossypium hirsutum* L.) and maize (*Zea mays* L.). Soil and Till. Res. 74: 161-168.

Netondo , G. W., J. C. Onyango & E. Beck, 2004. Sorghum and salinity : I. Response of growth, water relation, and

ion accumulation to NaCl salinity . Crop Science. 44:797-805.

Nun NB, Plakhine D, Joel D, Mayer A (2003) Changes in the activity of the alternative oxidase in Orobanche seeds during conditioning and their possible physiological function. Phytochemistry 64:235-241.

Rad J. S., Alfatemi M. H., Rad M.S., Sen D. J.. Phytochemical and Antimicrobial Evaluation of the Essential Oils and Antioxidant Activity of Aqueous Extracts from Flower and Stem of *Sinapis arvensis* L. American Journal of Advanced Drug Delivery. 2013. 1(1).1-10.

Rad J. S., Rad M. S., da Silva J.A. T. Effects of Exogenous Silicon on Cadmium Accumulation and Biological Responses of *Nigella sativa* L. (Black Cumin). Communications in Soil Science and Plant Analysis.In Press. 2014.

Rad J. S., Rad S. M., Miri A.. Regulation of the Expression of Nitrate Reductase genes in Leaves of Medical plant, *Foeniculum vulgare* by Different Nitrate Sources. International Journal of Agriculture and Crop Sciences. 2013/5-24/2911-2916.

Rad J.S., Rad M. S., da Silva J.A. T.. Evaluation of allelopathic effects of methanolic extracts from *Salicornia herbacea* seed and leaves on germination and seedling growth in vitro of two medicinal plants and two weeds. Environmental and Experimental Biology (2014) 12: 83–87.

Rad, J. S., Alfatemi S. M. H., Rad, M. S., Iriti M.. 2014. Free Radical Scavenging and Antioxidant Activities of Different Parts of *Nitraria schoberi* L. Journal of Biologically Active Products from Nature, 4(1): 44-51.

Rad, M. S. Rad, J. S. 2013. Effects of Abiotic Stress Conditions on Seed Germination and Seedling Growth of Medical Plant, Hyssop (*Hyssopus officinalis* L.). International Journal of Agriculture and Crop Sciences, 5(21): 2593-2597.

Senaratna, T., D. Touchel., E. Bumm & K. Dixon, 2000. Acetyl salicylic acid induces multiple stress tolerance in bean and tomato plants. Plant growth regulation. 30: 157-161.

Shah J (2003) The salicylic acid loop plant defense. Curr opin Plant Biol. 6: 365-371.

Smirnoff, N. & G.L.Wheeler , 2000. Ascorbic acid in Plants: biosynthesis and function, Critical reviews in plant sciences. 19: 267-290.

Smirnoff, N., 1996. The function and metabolism of ascorbic acid in plants, Annals of botany. 78: 661-669.

Tobe, K., Li, X & K.Omasa, 2000. Seed germination and radicle growth of a halophyte, *Kalidium capsicum* (Chenopodiaceae). Annals of botany. 85: 391-396.

Toker, C., S. Ulger., M. Karhan., H. Canci & O. Akdesir., N, Ertoy & M.I. Cagirgan, 2004. Comparison of some endogenous hormone levels in different parts of chickpea (*Cicer ariet*inum L.). Genet Resour Crop. 52, 233-237.

Uniyal, R.C and Nautiyal, A.R. 1998. Seed germination and seedling extension growth in (*Ougeinia dalbergioides* Benth.) under water and salinity stress. New Forests. 17,265–272.

Yuan SUN, Y., Y. Jian SUN., M. Tian wang, X. Yi LI., X. Guo & R.E. Hu, 2010. Effects of Seed Priming on Germination and Seedling Growth Under Water Stress in Rice. Acta Agronomica Sinica. 36(11): 1931-1940.

Zeng, Y.J., Wang, Y.R & J.M. Zhang, 2010. reduced seed germination due to water limitation a special survival strategy used by xerophytes in arid dunes. Journal of Arid environments. 74(4): 508-511.

 Table 1:

 Variance analysis of studied traits in Caper seeds under drought stress

Square means							
Seed vigor index	Seedling length	Shoot length	Rootl et length	Germinati on rate	Germina tion percentage	Freedo m degree	Sources of variation
116794.3**	11.24**	6.25 **	$0.772^{*}_{*}$	4.06**	481.6 **	9	Priming
3091782.91 **	480.49**	315.88**	17.82 <sup>*</sup>	105.19**	37984.9* *	4	Drought stress
12489.23**	0.92**	0.55**	0.091 <sup>*</sup>	0.54**	57.16**	36	Priming× Drought stress
1807.10	0.065	0.023	0.013	0.014	12.68	150	Error

\*\* Indicate significant difference at 1% probability level.

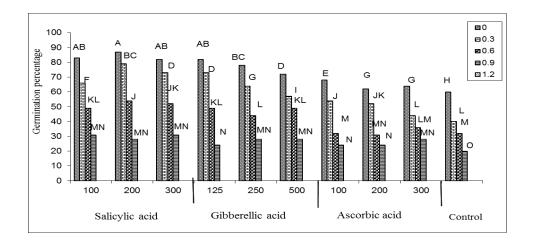


Figure 1. Effect of seed priming on germination percentage of C. Spinosa under different level of drought stress.

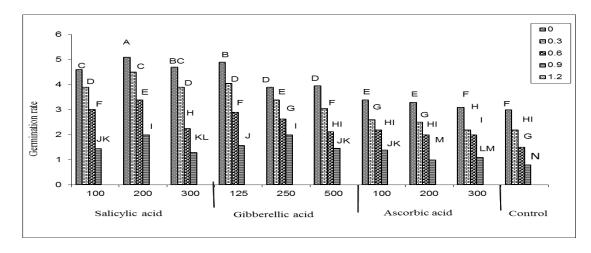


Figure 2. Effect of seed priming on germination rate of C. Spinosa under different level of drought stress.

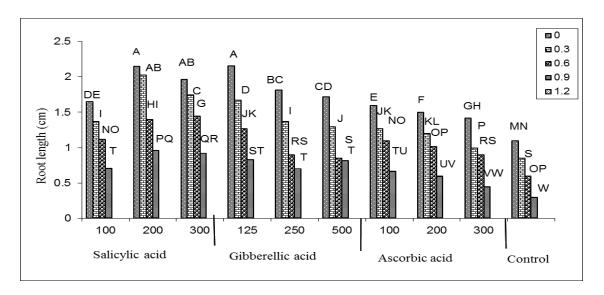


Figure 3. Effect of seed priming on root length of C. Spinosa under different level of drought stress.

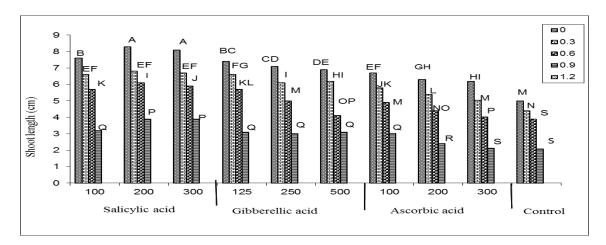


Figure 4. Effect of seed priming on shoot length of C. Spinosa under different level of drought stress.

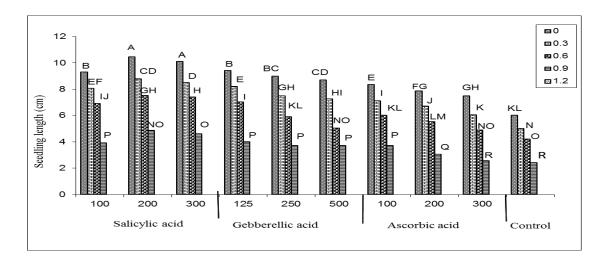


Figure 5. Effect of seed priming on seedling length of C. Spinosa under different level of drought stress.

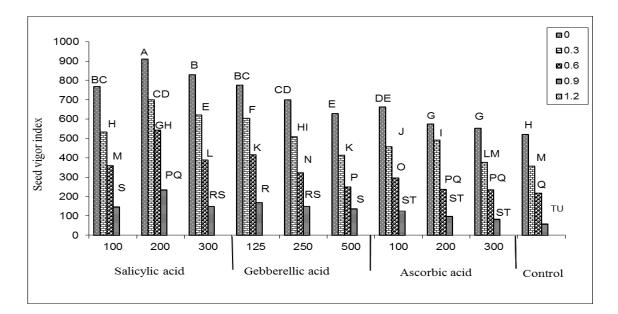


Figure 6. Effect of seed priming on seed vigor index of C. Spinosa under different level of drought stress.